Review Article

ISSN: 2349 – 4492



Asian Journal of Research in Biological and Pharmaceutical Sciences

Journal home page: www.ajrbps.com

https://doi.org/10.36673/AJRBPS.2020.v08.i03.A12



ROLE OF EPIGENETICS IN ETIOLOGY AND PREVENTION OF HUMAN DISEASES: A REVIEW BASED ON CURRENT EVIDENCES

Aditi Munmun Sengupta^{*1}, Diptendu Chatterjee², Debasis Das³, Salil Kumar Bhattacharya³, Rima Ghosh⁴

^{1*}C K Birla Hospitals, University of Calcutta, Harvard Medical School, Association Member, Kolkata, India.
²Department of Anthropology, Deputy Registrar of University of Calcutta, Kolkata, India.
³Department of Community Medicine, Medical College, Kolkata, India.
⁴Junior Research Fellow (UGC-NET), University of Calcutta, Kolkata, India.

ABSTRACT

In recent years, there has been an explosion of research on genetic causes of human diseases. Although genetic changes involve alteration of the genetic code, epigenetic changes are associated with external modifications to chromatin structure and gene expression without the alteration of the core DNA sequence. It is becoming increasingly evident that there are several disease-causing alterations in the epigenome. Thus, it is important to understand the etiology of human diseases with respect to epigenetic modifications. This review article focuses on the current advances in knowledge on human diseases that occur due to epigenetic dysregulation. We also highlight the role of epigenetics in diabetes regulation and in emerging viral infections. Understanding the significance of epigenetic changes and their role in disease development could help develop therapies targeting the epigenomic network of the cell.

KEYWORDS

DNA methylation, Histone modification and Epigenetic dysregulation.

Author for Correspondence:

Aditi Munmun Sengupta,

C K Birla Hospitals, University of Calcutta,

Harvard Medical School, Kolkata, India.

Email: sengupta2aditi@gmail.com

Available online: www.uptodateresearchpublication.com

INTRODUCTON

The term epigenetics was first introduced by Conrad Waddington in the early 1940s¹. Epigenetics involves the study of heritable changes in gene function that do not involve changes in the genetic code. Knowledge of epigenetics helps in understanding basic mechanisms where alterations of genes and gene products influence cell development. Epigenetic mechanisms directly influence gene expression at the transcriptional and translational levels and thus have the potential to

control overall gene expression. Epigenetic changes are known to regulate differentiation of pluripotent stem cells and determine the ultimate fate of the stem cell². Epigenetic regulation is involved in a spectrum of processes, large including morphogenesis, differentiation, imprinting, gene silencing, position effect, reprogramming and carcinogenesis. Several different epigenetic mechanisms exist in the cell, which include including methylation of DNA, post-translational modifications (acetylation, phosphorylation and sumoylation) of his tone proteins and small RNAmediated control of gene expression. Epigenetic modifications, and their subsequent effect on gene expression, are highly cell- and tissue-specific, enabling control of the gene expression profile in cells and tissues. These modifications are stable and are somatically heritable in dividing cells³. Thus, any aberration in the epigenetic state can result in altered tissue development and subsequent pathology. Recently, a wide range of human pathologies such as cancer, autoimmune disorders, diabetes, neurological disorders and interestingly, viral infections have been associated with de novo or inherited epigenetic changes^{4,5}. In this review, we examine the development of the study of epigenetics, and its role in healthy physiology and disease pathologies. Understanding the etiology, which is the detailed study of underlying mechanisms of disease development of these epigenetic changes, will help in better disease management and in developing effective therapy.

TYPES OF EPIGENETIC MODIFICATIONS

Following the discovery of the chromosome in 1879 by Ian Fleming^{6,7}, further studies, such as those by Thomas Morgan, helped to understand the genetic linkage between various chromosomes and X chromosome to understand sex determination in Drosophila. In the 1930s, HJ Muller carried out pioneering work to understand chromosomal rearrangements and their effect on Drosophila phenotype. This work was followed by research by Reik and Surani, who provided compelling imprinting evidence genomic and of the consequences on embryonic development⁸. Studies by Allfrey et al. in the 1960s established that histones post-translationally modified⁹. are

Extensive research following this pioneering discovery has shown that these modifications also alter gene expression patterns¹⁰. The most important of these modifications include histone 3 (H3) lysine 9 (K9) demethylation and lysine 27 (K27) trimethylation, which are involved in transition of heterochromatin to euchromatin and functional transcription of genes. With the advent of novel gene detection and sequencing technologies a wide array of epigenetic modifications have now been studied (Figure No.1), of which DNA methylation has been one of the most pioneering discoveries made in recent times. Many studies have since analyzed DNA methylation patterns and alteration of gene expression in various tissues. DNA methyltransferases are crucial in modifying methylation status of genes and play important roles in switching genes "on" and "off", leading to changes in gene expression. Studies on epigenetic regulation have been able to progress even further based on various microarray-based methods that have been developed. More recently, microRNAs (miRNA) and other non-protein coding RNAs (ncRNA) have been shown to regulate chromatin modification and thus gene expression¹¹. miRNAs regulate approximately 60% of genes in humans and slowly, but steadily, the expression of many miRNAs is being shown to be epigenetically regulated by histone modification or DNA methylation at specific loci¹².

- 1. DNA methylation: CH₃ (methyl group) is added by DNA methyl transferases (DNMT) to tag DNA and is associated with gene activation and repression.
- 2. Histone modifications: The binding of epigenetic factors to histone "tails" alters the extent to which DNA is wrapped around histones and the availability of genes in the DNA that may be activated or repressed.

Acetylation

COCH₃ (acetyl) group is added by histone acetyl transferases and is associated with gene activation.

Deacetylation

Acetyl group is removed by histone deacetyl (HDAC) transferases and is associated with gene repression.

EPIGENESIS AND **EPIGENETIC REGULATION IN NORMAL PHYSIOLOGY** Epigenesis plays an important role during development^{13,14}. Pluripotent stem cells in the embryo undergo differentiation to form tissuespecific oligo potent progenitor cells. Differentiation of pluripotent stem cells to their respective cell lineages is dependent upon the epigenetic changes in the genome that control the expression of specific genes. These epigenetic modifications alter the response of various transcriptional factors to hormones and other environmental cues. Thus, epigenesis plays an important role in overall determination of cellular fate and development of the whole organism.

The nature of epigenetic alterations has been understood by studying pluripotent stem cells¹⁵. Transition of adult somatic cells to pluripotent stem cells illustrates the major epigenetic changes that are involved in the whole process. Somatic cell programming during the transition involves a host of transcription factors, which include c-MYC, OCT-4, KLF4 and SOX2. The epigenetic landscape in pluripotent stem cells is maintained by genes such as OCT4 and NANOG. Expression of these two genes are silenced in somatic cells by DNA methylation at their promoters and repressive histone modifications such as H3K27 trimethylation (H3K27me3). The DNA methyl transferase DNMT3 also serves as a marker of reprogramming Reprogramming adult cells cells. of and establishment of pluripotent cells is marked by the establishment of epigenetic changes associated with the genome. The variety of the changes that are seen include altered levels of H3K4 and H3K27 trimethylation at signature gene promoters, activation of inactive X chromosome in female pluripotent cells, DNA hypomethylation of the heterochromatin repeats and maintenance of DNA methylation patterns at imprinted genes. The onset of these various events results in the final step of chromatin fiber reorganization and thus affects the overall elasticity and flexibility of chromatin. These various changes are important for appropriate functioning of pluripotent stem cells. Aberrations in any of these events results in induction of apoptotic pathways and cell death¹⁶.

ENVIRONMENTAL FACTORS GOVERNING EPIGENESIS

Epigenetic modifications are reversible. The genome is highly flexible and responds to a variety of environmental stimuli, such as nutrition and stress, as well as exposure to toxins and drugs. Food plays an important role in governing the overall growth and health of cells. Severe shortage of nutrition results in a lack of growth that could be the result of alteration of critical genes required for this. There is evidence that famines and a lack of nutritional food results in long term effects on the growth of children¹⁷. Poor nutrition also causes faulty imprinting of epigenetic signature sequences that results in development of neurological disorders and stress related disorders^{18,19}.

Folate, a water-soluble B vitamin, is one of the major nutritional components, which has been shown to affect epigenetic modifications²⁰. It is an important cofactor for many of the methyl transferases and thus plays an important role in DNA methylation. Consequently, folate deficiency been shown to be responsible has for hypomethylation of DNA and subsequent development of colorectal cancer²¹ and can also contribute to neural tube and crest defects during development²².

Exposure to toxic metals in the environment can also have an impact on the overall etiology of disease. Exposure to arsenic is known to be associated with altered DNA methylation levels, histone modification machinery, and miRNA expression, resulting in carcinogenesis and associated neurological defects²³. Prolonged use of certain drugs can also influence global epigenetic modifications. For instance, continuous use of oral contraceptive pills in women results in global hypomethylation of DNA²⁴. In addition, the use of procainamide and hydralazine modifies the nuclear architecture and development of anti-nuclear antibodies, resulting in drug-induced lupus²⁵.

Stress is another major contributing factor in development of disease. People with post-traumatic stress disorder have altered levels of DNA methylation and gene expression when compared with those who have not been stressed²⁶. Stress during pregnancy and perinatal stress has also been associated with the child developing neurological

Available online: www.uptodateresearchpublication.com

and psychiatric disorders²⁷. In addition, persistent exposure to pathogenic microorganisms results in modification of the genome and alteration in gene expression patterns²⁸. Many microbes are known to modify the epigenomic machinery, including the histone modifying enzymes, resulting in persistent infection of individuals. Indeed, inflammation has been proposed to be the common pathway of stress-related diseases, which could be attributed to epigenetic modifications^{29,30}.

Physical exercise has also been identified as an important factor that helps in controlling cancer development and progression^{31,32}. Physical exercise has been shown to up-regulate the expression of tumor suppressor genes, while down-regulating expression of oncogenes. Pathologies with dysregulated epigenetic modifications, which result in uncontrolled expression of several of the genes, can be regulated by physical activity³³.

HUMAN DISEASES CAUSED BY EPIGENETIC DYSREGULATION

There is increasing evidence that pathological epigenetic modifications are found in cancer, neurodegenerative diseases, and ageing (Figure No.2).

DNA and histone proteins are associated together by electrostatic interactions. DNA wraps around the histone proteins whose tails protrude from the structure. The epigenome is a series of chemical modifications that occur in DNA or specific amino acids in histone proteins that DNA is wrapped around. These act as markers that determine whether genes are active or inactive at certain times. These epigenetic changes are responsible for human diseases.

CANCER

Cancer is one of the most well studied pathologies where aberrant epigenetic modifications are involved in disease activity. Gross changes in DNA methylation patterns are seen in cancer cells when compared to non-cancerous cells. During tumorigenesis, both genome-wide and regional epigenetic changes occur, which drive tumor progression^{34,35}. Hypermethylation of various promoter regions or repressive histone signatures on promoters result in the inactivation of tumor

Available online: www.uptodateresearchpublication.com

suppressor genes³⁶. Hypermethylation of promoters, particularly in CpG island regions, for genes that support cell survival, such as APC, RASSF1A and TP53, is commonly observed in cancer cells. In fact, most cancers have distinct epigenetic alteration patterns. Non-small cell lung cancer exhibits levels of elevated DNMT, global deacetylated H3 and trimethylated H3K9^{37,38,39}. DNMT1 and histone deacetylase 1 (HDAC1) are up-regulated during progression of pancreatic cancers in a malignancydependent manner and consequently deregulate genes that control the hallmarks of cancer proliferation, survival, angiogenesis, invasion and metastasis^{40,41}. In colorectal cancer. global hypomethylation contributes to activation of various oncogenes, whereas hypermethylation in CpG islands promote tumorigenesis by silencing tumor suppressor genes. Multiple versions of translocation methylcytosine dioxygenase (TET1), an important DNA methylation regulator, were shown to be down-regulated in the early phases of tumorigenesis⁴². Histone modifications including loss of H3K20 trimethylation and di- and trimethylation of H3K4, H3K9 and H3K27 have been well documented in the pathogenesis of colorectal cancer⁴³.

AUTOIMMUNE DISEASES

Autoimmune diseases are another major class of diseases where epigenetic changes make a contribution to pathology 44 . significant In inflammation, DNA methylation contributes to T cell activation and function⁴⁵. Dysregulated T cell functions are important in driving autoimmunity⁴⁶. Global hypomethylation of DNA is observed in T from patients with systemic cells lupus erythematosus or rheumatoid arthritis⁴⁷. In these patients, regional hypermethylation of promoters associated with HDAC1 and HDAC2 is observed⁴⁸. Furthermore, the balance between HDACs and HATs is disturbed, resulting in hyperacetylation of histones H3 and H4. In osteoarthritis, regional H3K9 hypomethylation is associated with the pathophysiology⁴⁹. In patients with sclerosis, hypomethylation at H3K27 me3 has been observed in CD4+ T cells as well in the white matter of the central nervous system⁵⁰.

NEURODEGENERATIVE DISEASES

Neurodegenerative disorders have a distinct epigenomic imprint which is different from healthy individuals. In both Alzheimer's disease (AD) and schizophrenia, reduced DNA methylation occurs at promoter regions for genes expressed in the central nervous system. In addition to methylation at the 5th position of the pyrimidine ring of cytosine (5mC), hydroxy-modified 5mC (5hmc) is present at high levels in the healthy brain and central nervous system⁵¹. Reductions in levels of 5mC and 5hmC and also in levels of DNMT have been observed in AD^{52,53}. However, contrasting data has suggested that an increase in these levels was observed in the frontal cortex of AD patients⁵⁴. This indicates that epigenetic modifications of gene expression in the nervous system is highly temporal and further study is warranted to fully understand the epigenetic program that underlies a healthy conditions. Decreased methylation is observed in the Alu repeat sequences of the genome in AD patients and this serves as one of the indicative features for prognosis of the disease⁵⁵. Dementia and AD are also associated with alteredrhythmic methylation cycles of circadian rhythm protein BMAL₁ promoter resulting in rapid BMAL₁ degradation^{56, 57}. In frontotemporal dementia, hypermethylation of the promoter and potential progranulin (GRN)hypermethylation of chromosome 9 open reading frame 72 (C9ORF72) promoter has been observed as a genetic mechanism involved in the pathology of this disease⁵⁸.

Abnormal methylation and global DNA hypomethylation are associated with autism and related disorders. Hypermethylation at the promoter region of the FMR1 gene was found to be responsible for Fragile X syndrome, which may cause autism spectrum disorder⁵⁹. Schizophrenia is associated with large number of changes mediated by epigenetic interactions⁶⁰. The hypermethylation of sex-determining region Y-box containing gene 10 (SOX10) promoter in the brain leads to downregulation of SOX10 expression, H3K27me3 hypomethylation at the glutamic acid decarboxylase 67 (GAD1) promoter, DNA hypomethylation at the WDR18 gene, and elevated expression of DNMT1 and all these changes have been associated with schizophrenia⁶¹.

Available online: www.uptodateresearchpublication.com

DIABETES

According to statistics from WHO 2020, diabetes mellitus (DM) affects approximately 415 million people worldwide, of which 46% are undiagnosed and by 2040, 642 million will be affected. The most common types of DM present in the population are type 1 diabetes mellitus (T1DM) and type 2 diabetes mellitus (T2DM). T1DM is an autoimmune condition where pancreatic β -cells are destroyed, whereas T2DM is a metabolic condition that is caused by a shortage of insulin production by pancreatic cells. T2DM is the prevalent form of DM in the population and accounts for more than 85% of existing cases. Environmental factors, especially obesity and viral infections (in the case of T1DM), play a major role in the etiology of both types of diabetes^{62,63,64}. In general, DM is associated with cardiovascular pathologies, which stem from hyperglycemia, a common factor in both types of DM. Epigenetic changes seen in diabetic nephropathy, such as retinopathy, neuropathy and impaired wound healing have been extensively studied to understand the etiology of the disease^{65,66} (Figure No.3). Pathologies, such as diabetic retinopathy, have been shown to progress despite resolution of glycemic control, suggesting epigenetic control or metabolic memory^{67,68}. Studies mouse models of diabetic retinopathy in demonstrate increased acetylation of H3K9 and trimethylation of H4K20. increased These alterations led to decreased expression of superoxide dismutase (SOD2) and retinal MnSOD, resulting in on set of diabetic retinopathy. The histone deacetylase SIRT1 is also down-regulated in hyperglycemic rats. SIRT1 is a target of miR-23b-3p, which is up-regulated in retinal endothelial cells, even in the absence of hyperglycemia⁶⁹. Other noncoding RNAs have also been implicated in the pathology of retinopathy and nephropathy. miR-200b and miR-146a have been shown to play an important role in diabetic retinopathy⁷⁰, whereas miR-29c, miR-21, miR-200 and miR-192 have been nephropathy^{71,72}. implicated in diabetic either Hyperglycemia results in DNA hypomethylation or hypermethylation in rat models of diabetes in a tissue-specific manner^{73,74}. Damage to mitochondria is a distinct feature present in both diabetic retinopathy and nephropathy⁷⁵. Diabetes

increases mitochondrial DNA (mtDNA) damage, leading to reduction in abundance of healthy mitochondria. Epigenetic modifications due to hyperglycemia affect enzymes important in mitochondrial homeostasis and thus enhance mitochondrial damage. The transport of mitochondrial transcriptional factor A (TFAM), which is essential for mtDNA replication, is also blocked in diabetes mellitus patients because of epigenetic-like modifications^{76, 77}.

Diabetes induces hyperglycemia that activates the reactive oxygen species (ROS) pathways causing oxidative stress. This is turn regulates epigenetic factors such as histone methyl transferases (HMTs), acetvl transferases (HATs), histone histone deacetylases (HDACs) and DNA methyltransferases (DNMTs) to enable histone post translational modifications (HPTMs) and DNA methylation (DNA-me) leading to changes in gene expression. Persistence of these epigenetic changes may help to develop metabolic memory, which can further aggravate the oxidative stress and enable continued development of complications. Mitochondrial homeostasis is also affected by alteration in gene expression, thus enhancing mitochondrial damage resulting in chronic complications.

VIRAL INFECTIONS

Emerging viral infections pose a major threat to global public health. In 2019, COVID-19 developed into a global pandemic, which has persisted well into 2020⁷⁸. The causative virus, severe acute respiratory syndrome-coronavirus 2 (SARS-CoV-2) belongs to the coronavirus family, a large family of viruses that are common in humans and animals. Similar to SARS-CoV and MERS-CoV (Middle East respiratory syndrome-coronavirus), SARS-CoV-2 infects the respiratory system and has a zoonotic origin. SARS-CoV-2 may cause mild to moderate upper respiratory infections but can progress to more severe symptoms, such as bronchitis and pneumonia. A striking difference between SARS-CoV-2 and SARS-CoV and MERS-CoV is the ability to cause silent infections early on. COVID-19 can range from silent and barely symptomatic infections to critical illness^{79, 80}. Viruses such as COVID-19 can spread and mutate very quickly in humans. Epigenetic studies may

Available online: www.uptodateresearchpublication.com

provide essential information to understand COVID-19 infections. In viral-host interactions, DNA-RNA methylation, chromatin remodeling and histone modifications are known to regulate and remodel host expression patterns⁸¹. RNA-type viruses, such as SARS-CoV-2, exhibit strong associations with internal RNA modifications. For instance, m6A, m6Am and 2'-O-me play important roles in the viral lifecycle^{82, 83}. These can affect viral structure and replication, as well as the innate immune response and innate immune regulatory pathways. Viruses, such as those from the coronavirus family and influenza, are not able to make genetic modifications to their hosts; however, they can alter the epigenome, allowing them to evade the host immune response and successfully spread infection. Epigenetic mechanisms were identified as a common avenue used by MERS-CoV and H₅N₁ in "switching off" host immune responses⁸⁴. Although DNA methylation was the primary mechanism used to suppress production of antigen presentation molecules in MERS-CoV infections, both DNA methylation and histone methylation were involved in reducing the immune response to H_5N_1 . The influenza-A virus (H_3N_2) inhibits the immune response of the host by producing a protein, NS1, that mimics the tail of the histone and can interact with the host transcription complex and block antiviral gene expression in the host⁸⁵.

Studies are now ongoing to explore whether infection with the SARS-CoV-2 virus induces longlasting immune response. A failure to induce longterm immune memory would impose serious challenges for vaccine development. Reinfection may also occur if the virus mutates enough to evade immune recognition⁸⁶. SARS-CoV-2 particles are pleomorphic (i.e., they lack a defined structure); however, under acryoelectron microscopy, a helical nucleic acid structure is apparent. The SARS-CoV-2 genome is encoded in an enveloped positive-sense unsegmented single-strand of RNA. This RNA is directly read by ribosomes of the host cell⁸⁷. In Figure No.4, the possible immune mechanisms present COVID-19 have been explained in detail.

1.) The virus enters the cell via interaction of the viral spike protein and receptors in the host cell-membrane. On entry into the cell, the viral genome

triggers immune response signaling in the infected cell 2.) The SARS-CoV-2 genome possesses ~14 open reading frames (ORFs) encoding approximately 27 proteins. The longest ORF is located at the 5' end of the genome and encodes 15 nonstructural proteins collectively involved in virus replication and possibly immune invasion. The 3' end of the genome encodes structural and accessory proteins. Accessory proteins are not required for virus replication or other known functions. Genbank reference genome (NC_045512.2.3) Biological changes occur in the infected cell and create interconnected double-membrane vesicles that form the replication and transcription complexes (RTCs) during SARS-CoV-2 replication. RTCs are potential antiviral targets. Immune interactions, particularly the innate immune system, are likely the driving force behind the pathogenesis of SARS-CoV-2 virus in human hosts.

The active replication and release of SARS-CoV-2 causes the host cell to undergo pyroptosis and release damage-associated molecules such as ATP, nucleic acids and ASC oligomers. These are recognized by epithelial cells and alveolar macrophages, triggering the generation of proinflammatory cytokines and chemokines, attracting monocytes, macrophages and Tcells to the site of infection and promoting further inflammation and establishing a proinflammatory feedback loop⁸⁸. In an immune-compromised individual, this may lead to further accumulation of immune cells in the lungs, causing overproduction of pro-inflammatory cytokines, which eventually damages the lung infrastructure. The resulting cytokine storm circulates to other organs, leading to multi-organ damage. In addition, non-neutralizing antibodies produced by B-cells may enhance SARS-CoV-2 infection through antibody-dependent enhancement further exacerbating organ damage. ARDS is the primary feature of SARS-CoV-2 infection characterized by difficulty in breathing and low blood oxygen level^{86,89}.

The longevity of the antibody response to SARS-CoV-2 remains unknown. Most persons infected with SARS-CoV-2 display an antibody response 10 to 21 days after infection. However, detection in mild cases can take a longer time (four weeks or more), and in a small number of cases antibodies

Available online: www.uptodateresearchpublication.com

(i.e., IgM and IgG) are not detected at all within the time scale of the study. Based on the currently available data, the IgM and IgG antibodies to SARS-CoV-2 develop within 6 to 15 days post disease onset^{90,91}. The median seroconversion time for total antibodies, IgM and then IgG is day-11, day-14 dav-12 and post-symptom onset. respectively. The presence of antibodies was detected in <0.40% among patients within 1 week from disease onset, and rapidly increased to 100% (total antibodies), 94.3% (IgM) and 79.8% (IgG) from day-15 after onset⁹⁰. T cell responses against the SARS-CoV-2 spike protein correlate well with IgG and IgA antibody titers in COVID-19 patients, which has important implications for vaccine design and long-term immune response^{92,93}. Longitudinal serological studies that follow patient immunity over an extended period of time would be required to study the duration of immunity⁹⁴.

Two recent literatures^{95,96} identified the importance of methylation pattern of the gene encoding for angiotensin converting enzyme 2 (ACE2), which is the virus receptor on host lung epithelial cells for SARS-CoV-2. It has been shown that the production rate of ACE-2 enzyme by its gene is under epigenetic control⁹⁷. The expression of ACE2 and interferon gene depends on the methylation rate of the CpG islands in the DNA promoter sequence, which has influence on key inflammatory and immune responses in viral infections⁹⁸. Susceptible individuals, especially men, the elderly and the smokers show a hypomethylation pattern of the ACE2 and interferon genes, whereas women, children. and nonsmokers show DNA hypermethylation and lower expression of ACE2 and interferon proteins⁹⁹⁻¹⁰². High expression of interferon genes has been characterized by cytokine storm in severe SLE¹⁰³ and a cytokine storm is characteristic of SARS-CoV-2 infection¹⁰⁴. The common characteristics in Lupus and SARS-CoV-2 infection are due to increased oxidative stress and DNA demethylation of ACE2. Hypomethylation and over expression of ACE-2 in T-cells will facilitate T cell viral infection and viral dissemination resulting in increased severity of COVID-19¹⁰⁵. The example of cytokine storm in lupus may lead to the hypothesis that interferonregulated genes and other inflammatory cytokine

genes are hypomethylated and thereby are epigenetically primed for transcription upon interferon exposure resulting from the viral immune response. The epigenetic priming might increase the possibility of cytokine storm in susceptible patients. We can hence get an idea to the fact that that epigenetics may play a role in delayed immune response to COVID-19 infection or ongoing inflammation after infection has been resolved¹⁰⁶.

LEVERAGING EPIGENETICS FOR ETIOLOGY AND DISEASE PREVENTION

Epigenomic changes are recognized as contributing to the etiology of several diseases. Thus, understanding the distribution of epigenomic markers is important in assessing the phenotype and nature of the disease. Epigenetic epidemiology deals with the relationships between the epigenomic code and disease development in a large population. The most common epigenomic modification is DNA methylation. Systematic analysis of cancer cells by Feinberg et al. Demonstrated large-scale deletions in heterochromatin and DNA hypomethylation over a period in several cancers. Large organized chromatin lysine modifications (LOCKs) and lamina associated domains (LADs) were further analyzed for DNA methylation changes^{107,108}. Large regions of DNA, known as super-enhancers, are hypomethylated in LOCKs and LADs and this hypomethylation is preserved during further divisions, resulting in loss of expression of many of the genes in those genomic regions¹⁰⁹. Differentially methylated regions are hallmark features of several cancers. The identification of epigenetic markers would help these in understanding the significance and tumor progression in different tissues¹¹⁰.

Persistent exposure of cells to hyperglycemia results in epigenetic changes in the genome which are irreversible. Large-scale clinical trials have shown that diabetic patients continue to develop complications despite regaining control of blood sugar levels¹¹¹, which has been attributed to cells acquiring metabolic memory of previous hyperglycemic exposure to the cells¹¹². Metabolic memory leads to a distinct epigenetic imprint that has been observed in many studies. These have shown that decrease of trimethylation at histone 3

Available online: www.uptodateresearchpublication.com

lysine 9 serves as marker in many cases, leading to enhanced inflammation in diabetes^{113,114}. This decrease of trimethylation results in suppressed expression of many of the genes such as the H3K9me3 methyltransferase (Suv39h1) and increased expression of the reactive oxygen species $(p66Shc)^{115}$. (ROS)-regulator The lvsine methyltransferase SET7 is another epigenetic marker that is functionally altered in diabetic patients and helps to sustain metabolic memory, leading to vascular dysfunction¹¹⁶. Many smallmolecule inhibitors of epigenetic mediators are currently in use for treatment of diabetes. For instance, trichostatin A, suberoylanilide hydroxamic acid, MS275 and valproic acid are histone deacetylase inhibitors, whereas anacardic acid and garcinol are histone acetyl transferase inhibitors. HDAC inhibitors help in enhancement of insulin secretion, increase in insulin sensitivity, expression of anti-inflammatory genes and induction of beta cell proliferation and regeneration¹¹⁷. In addition, locked nucleic acid and antisense nucleotides against miRNAs are known to increase insulin sensitivity of cells.

Currently, we are facing a highly prolific viral outbreak. A lesson from the recent pandemic of SARS-CoV-2 is that vaccine development may take months to years. In the meantime, is it possible to influence the epigenetic regulation of multiple genes with natural interventions? The rapid dissemination of SARS-CoV-2 is a reminder of how future epigenetic studies can expand upon what we already know about these types of viruses. Specific aspects of the anti-viral response and dysregulation can be explained by epigenetics. Although epigenomic studies have found significance in understanding the progression of the disease, there is a need for more therapies based on epigenetic modifications. Manipulations of these epigenetic changes could form the basis of a therapeutic methodology by controlling viral infections and developing effective broad-spectrum antivirals¹¹⁸. A combination of genomic and epigenomic methods is required to understand the intricate changes associated with pathologies.

New advances in technology for studying the epigenetic landscape at a genome-wide scale and in a sequence-specific manner will therefore be

required to address diseases, especially rapidly COVID-19⁸⁰. emerging diseases. such as Methylation quantitative trait loci (meQTL) is one of the techniques that has been developed to understand the epigenomic changes and subsequent alterations in the genome, which could predict for the overall health of the tissue being analyzed¹¹⁰. Epigenetic study has also proved useful in developing biomaterial-based stem cell growth models. Pluripotent stem cells isolated from various tissues can be isolated, cultured in vitro and differentiated into defined cell types using a transcription combination of factors and environmental cues. The exposure of these cells in hydrogel models helps understand the various interactions that decide the epigenetic changes and the ultimate cell fate¹¹⁹. These systems are increasingly being used to model and understand genetic-epigenetic crosstalk in different the pathologies^{120,121}. They also have considerable potential to aid research with developing therapeutic interventions.



Figure No.2: Human diseases caused by epigenetic dysregulation

Available online: www.uptodateresearchpublication.com



Figure No.4: The predicted antiviral immune mechanism of SARS-CoV-2

CONCLUSION

Epigenomic modifications have distinct roles to play in maintenance of stem cell pluripotency, and distinct epigenetic signatures also control tissuespecific gene expression. Understanding these epigenomic changes, both at the global and regional levels, is vital to understanding their effect on pathological conditions. Detailed study of epigenomic changes in conjunction with genomic changes is required to gain insight into the molecular mechanisms underlying human disease.

Available online: www.uptodateresearchpublication.com

While advanced technology has aided research in the field, our understanding is impaired by the lack of high-throughput tools to study the crosstalk between genetic and epigenetic changes. Comprehensive analysis for epidemiological epigenetics needs to be done for each disease and consensus epigenomic changes need to be evaluated. Based on these observations, evaluation strategies for therapies can then be actively pursued.

ACKNOWLEDGEMENT

Special acknowledgement to Prof. Arup Ratan Bandopadhyay, Head of the Department of Anthropology, University of Calcutta for his able guidance and Prof. Britt Glaunsinger, University of California Berkley for helping in the concept development.

FINANCIAL SUPPORT AND SPONSORSHIP

Sri Sarosij Ray Memorial Research support fund.

CONFLICT OF INTEREST

We declare that we have no conflict of interest.

BIBLIOGRAPHY

- 1. Berger S L, Kouzarides T, Shiekhattar R, Shilatifard A. An operational definition of epigenetics, *Genes and Development*, 23(7), 2009, 781-783.
- Lu Q, Qiu X, Hu N, Wen H, Su Y, Richardson B C. Epigenetics, disease and therapeutic interventions, *Ageing Research Reviews*, 5(4), 2006, 449-467.
- 3. Zhang J, Gao Q, Li P, *et al.* S phase-dependent interaction with DNMT1 dictates the role of UHRF1 but not UHRF2 in DNA methylation maintenance, *Cell Research*, 21(12), 2011, 1723-1739.
- Abdolmaleky H M, Zhou J R, Thiagalingam S, Smith C L. Epigenetic and pharmacoepigenomic studies of major psychoses and potentials for therapeutics, *Pharmaco*, 9(12), 2008, 1809-1823.
- 5. Lu C, Thompson C B. Metabolic regulation of epigenetics, *Cell Metab*, 16(1), 2012, 9-17.
- 6. Felsenfeld G. A brief history of epigenetics, *Cold Spring Harb Perspect Biol*, 6(1), 2014, a018200.
- 7. Holliday R. Epigenetics: A historical overview, *Epigenetics*, 1(2), 2006, 76-80.
- 8. Reik W, Surani M A. Genomic imprinting and embryonal tumours, *Nature*, 338(6211), 1989, 112-113.
- 9. Allfrey V G, Faulkner R, Mirsky A E. Acetylation and methylation of histones and their possible role in the regulation of RNA synthesis, *Proc Natl Acad Sci United States*, 51(5), 1964, 786-794.

Available online: www.uptodateresearchpublication.com

- 10. Feinberg A P. The epigenetic basis of common human disease, *Trans Am Clin Climatol Assoc*, 124, 2013, 84-93.
- 11. Yao Q, Chen Y, Zhou X. The roles of micro RNAs in epigenetic regulation, *Curr Opin Chem Biol*, 51, 2019, 11-17.
- 12. Friedman R C, Farh K K H, Burge C B, Bartel D P. Most mammalian mRNAs are conserved targets of micro RNAs, *Genome Res*, 19(1), 2009, 92-105.
- 13. Wagers A J, Christensen J L, Weissman I L, Cell fate determination from stem cells, *Gene Ther*, 9(10), 2002, 606-612.
- Ichida J K, Kiskinis E and Eggan K. Shushing down the epigenetic landscape towards stem cell differentiation, *Development*, 137(15), 2010, 2455-2460.
- 15. Watanabe A, Yamada Y, Yamanaka S. Epigenetic regulation in pluripotent stem cells: A key to breaking the epigenetic barrier, *Philos Trans R Soc B Biol Sci*, 368(1609), 2013, 20120386.
- 16. Djuric U, Ellis J. Epigenetics of induced pluripotency, the seven-headed dragon, *Stem Cell Res Ther*, 1(1), 2010, 3.
- Gomez-Verjan J C, Barrera-Vazquez O S, Garcia-Velazquez L, Samper-Ternent R, Arroyo P. Epigenetic variations due to nutritional status in early-life and its later impact on aging and disease, *Clin Genet*, 2020.
- Van Eijk K R. Quantitative studies of dna methylation and gene expression in neuropsychiatric traits, *Utrecht University Repository*, 2014, 1-181.
- 19. Mehta D, Klengel T, Conneely K N *et al.* Childhood maltreatment is associated with distinct genomic and epigenetic profiles in posttraumatic stress disorder, *Proc Natl Acad Sci U S A*, 110(20), 2013, 8302-8307.
- 20. Kim K chol, Friso S, Choi S W. DNA methylation, an epigenetic mechanism connecting folate to healthy embryonic development and aging, *J Nutr Biochem*, 20(12), 2009, 917-926.
- 21. Kim Y I. Nutritional Epigenetics: Impact of folate deficiency on DNA methylation and colon cancer susceptibility, *The Journal of Nutrition*, 135(11), 2005, 2703-2709
- July September

- 22. Alata Jimenez N, Torres Perez S A, Sanchez-Vasquez E, Fernandino J I, Strobl-Mazzulla P H. Folate deficiency prevents neural crest fate by disturbing the epigenetic Sox2 repression on the dorsal neural tube, *Dev Biol*, 444(1), 2018, S193-S201.
- 23. Ren X, Mchale C M, Skibola C F, Smith A H, Smith M T, Zhang L. An emerging role for epigenetic dysregulation in arsenic toxicity and carcinogenesis, *Environ Health Perspect*, 119(1), 2011, 11-19.
- 24. Greer J M, Mccombe P A. The role of epigenetic mechanisms and processes in autoimmune disorders, *Biol Targets Ther*, 6, 2012, 307-327.
- 25. Borchers A T, Keen C L, Gershwin M E. Druginduced lupus, *In: Annals of the New York Academy of Sciences, Blackwell Publishing Inc*, 1108, 2007, 166-182.
- 26. Zawia N H, Lahiri D K, Cardozo-Pelaez F. Epigenetics, oxidative stress and Alzheimer disease, *Free Radic Biol Med*, 46(9), 2009, 1241-1249.
- 27. Babenko O, Kovalchuk I, Metz GAS. Stressinduced perinatal and transgenerational epigenetic programming of brain development and mental health, *Neurosci Biobehav Rev*, 48, 2015, 70-91.
- 28. Hamon M A, Batsche E, Regnault B *et al.* Histone modifications induced by a family of bacterial toxins, *Proc Natl Acad Sci, USA*, 104(33), 2007, 13467-13472.
- 29. Taccagni G L, Carlucci M, Sironi M, Cantaboni A, Di Carlo V. Duodenal somatostatinoma with psammoma bodies: An immunohistochemical and ultrastructural study, *Am J Gastroenterol*, 81(1), 1986, 33-37.
- 30. Gonzalez-Jaramillo V, Portilla-Fernandez E, Glisic M *et al.* Epigenetics and inflammatory markers: A systematic review of the current evidence, *Hindawi Int J Inflamm*, 2019, Article ID: 6273680, 2019, 14.
- 31. Zeng H, Irwin M L, Lu L *et al.* Physical activity and breast cancer survival: An epigenetic link through reduced methylation of a tumor suppressor gene L3MBTL1, *Breast Cancer Res Treat*, 133(1), 2012, 127-135.
- 32. Mctiernan A, Friedenreich C M, Katzmarzyk P

 $\label{eq:available} Available \ on line: www.uptodate research publication.com$

T *et al.* Physical Activity in Cancer Prevention and Survival: A systematic review, *Med Sci Sports Exerc*, 51(6), 2019, 1252-1261.

- 33. Ferioli M, Zauli G, Maiorano P, Milani D, Mirandola P, Neri L M. Role of physical exercise in the regulation of epigenetic mechanisms in inflammation, cancer, neurodegenerative diseases and aging process, J *Cell Physiol*, 234(9), 2019, 14852-14864.
- 34. Baylin S B, Jones P A. Epigenetic determinants of cancer, *Cold Spring Harb Perspect Biol*, 8(9), 2016, e019505.
- 35. Esteller M. Non-coding RNAs in human disease, *Nat Rev Genet*, 12(12), 2011, 861-874.
- 36. Fraga M F, Ballestar E, Villar-Garea A *et al.* Loss of acetylation at Lys16 and trimethylation at Lys20 of histone H4 is a common hallmark of human cancer, *Nat Genet*, 37(4), 2005, 391-400.
- 37. Li J, Tao X, Shen J *et al.* The molecular landscape of histone lysine methyltransferases and demethylases in non-small cell lung cancer, *Int J Med Sci*, 6(7), 2019, 1922-1930.
- 38. Brock M V, Hooker C M, Ota-Machida E *et al.* DNA methylation markers and early recurrence in stage I lung cancer, *N Engl J Med*, 358(11), 2008, 1118-1128.
- 39. Damiani L A, Yingling C M, Leng S, Romo P E, Nakamura J, Belinsky S A. Carcinogeninduced gene promoter hypermethylation is mediated by DNMT1 and causal for transformation of immortalized bronchial epithelial cells, *Cancer Res*, 68(21), 2008, 9005-9014.
- 40. Wang W, Gao J, Man XH, Li Z S, Gong Y F. Significance of DNA methyltransferase-1 and histone deacetylase-1 in pancreatic cancer, *Oncol Rep*, 21(6), 2009, 1439-1447.
- 41. Neureiter D, Jager T, Ocker M, Kiesslich T. Epigenetics and pancreatic cancer: Pathophysiology and novel treatment aspects, *World J Gastroenterol*, 20(24), 2014, 7830-7848.
- 42. Neri F, Dettori D, Incarnato D *et al.* TET1 is a tumour suppressor that inhibits colon cancer growth by derepressing inhibitors of the WNT pathway, *Oncogene*, 34(32), 2015, 4168-4176.
- 43. Gargalionis A N, Piperi C, Adamopoulos C, Papavassiliou A G. Histone modifications as a
- July September

pathogenic mechanism of colorectal tumorigenesis, *Int J Biochem Cell Biol*, 44(8), 2012, 1276-1289.

- 44. Mazzone R, Zwergel C, Artico M *et al.* The emerging role of epigenetics in human autoimmune disorders, *Clin Epigenetics*, 11(1), 2019, 34.
- 45. Scharer C D, Barwick B G, Youngblood B A, Ahmed R, Boss J M. Global DNA methylation remodeling accompanies CD8 T cell effector function, *J Immunol*, 191(6), 2013, 3419-3429.
- 46. Goverman J. Autoimmune T cell responses in the central nervous system, *Nat Rev Immunol*, 9(6), 2009, 393-407.
- 47. Richardson B, Scheinbart L, Strahler J, Gross L, Hanash S, Johnson M. Evidence for impaired T cell DNA methylation in systemic lupus erythematosus and rheumatoid arthritis, *Arthritis Rheum*, 33(11), 1990, 1665-1673.
- 48. Huber L C, Brock M, Hemmatazad H *et al.* Histone deacetylase/acetylase activity in total synovial tissue derived from rheumatoid arthritis and osteoarthritis patients, *Arthritis Rheum*, 56(4), 2007, 1087-1093.
- 49. Ukita M, Matsushita K, Tamura M, Yamaguchi T. Histone H3K9 methylation is involved in temporomandibular joint osteoarthritis, *Int J Mol Med*, 45(2), 2020, 607-614.
- Wang Q, Xiao Y, Shi Y *et al.* Overexpression of JMJD3 may contribute to demethylation of H3K27me3 in CD4+ T cells from patients with systemic sclerosis, *Clin Immunol*, 161(2), 2015, 396-399.
- 51. Shi D Q, Ali I, Tang J, Yang W C. New insights into 5hmC DNA modification: Generation, distribution and function, *Front Genet*, 8, 2017, 100.
- 52. Bihaqi S W, Schumacher A, Maloney B, K. Lahiri D, H. Zawia N. Do epigenetic pathways initiate Late Onset Alzheimer Disease (LOAD): towards a new paradigm, *Curr Alzheimer Res*, 9(5), 2012, 574-588.
- 53. Chouliaras L, Mastroeni D, Delvaux E *et al.* Consistent decrease in global DNA methylation and hydroxymethylation in the hippocampus of Alzheimer's disease patients, *Neurobiol Aging*, 34(9), 2013, 2091-2099.
- 54. Rao J S, Keleshian V L, Klein S, Rapoport S I.

Available online: www.uptodateresearchpublication.com

Epigenetic modifications in frontal cortex from Alzheimer's disease and bipolar disorder patients, *Transl Psychiatry*, 2(7), 2012, e132.

- 55. Bollati V, Galimberti D, Pergoli L *et al.* DNA methylation in repetitive elements and Alzheimer disease, *Brain Behav Immun*, 25(6), 2011, 1078-1083.
- 56. Cronin P, McCarthy M J, Lim A S P *et al.* Circadian alterations during early stages of Alzheimer's disease are associated with aberrant cycles of DNA methylation in BMAL1, *Alzheimer's Dement*, 13(6), 2017, 689-700.
- 57. Tomita T, Onishi Y. Epigenetic modulation of circadian rhythms: Bmal1 gene regulation, *Chromatin and Epigenetics, Intech Open,* Chapter 6, 2020, 1-20.
- 58. Fenoglio C, Scarpini E, Serpente M, Galimberti D. Role of genetics and epigenetics in the pathogenesis of Alzheimer's disease and frontotemporal dementia, *J Alzheimer's Dis*, 62(3), 2018, 913-932.
- 59. Kraan C M, Godler D E, Amor D J. Epigenetics of fragile X syndrome and fragile X-related disorders, *Dev Med Child Neurol*, 61(2), 2019, 121-127.
- 60. Landgrave-Gomez J, Mercado-Gomez O, Guevara-Guzman R. Epigenetic mechanisms in neurological and neurodegenerative diseases, *Front Cell Neurosci*, 9(56), 2015, 1-11.
- 61. Iwamoto K, Bundo M, Yamada K *et al.* DNA methylation status of SOX10 correlates with its downregulation and oligodendrocyte dysfunction in schizophrenia, *J Neurosci*, 25(22), 2005, 5376-5381.
- 62. Altobelli E, Petrocelli R, Verrotti A, Chiarelli F, Marziliano C. Genetic and environmental factors affect the onset of type 1 diabetes mellitus, *Pediatr Diabetes*, 17(8), 2016, 559-566.
- 63. Liu Y, Lou X. Type 2 diabetes mellitus-related environmental factors and the gut microbiota: Emerging evidence and challenges, *Clinics*, 75, 2020, e1277.
- 64. Rewers M, Ludvigsson J. Environmental risk factors for type 1 diabetes, *Lancet*, 387(10035), 2016, 2340-2348.
- 65. Cerna M. Epigenetic regulation in etiology of type 1 diabetes mellitus, *Int J Mol Sci*, 21(1),
- July September

2020, 36.

- 66. Parrillo L, Spinelli R, Nicolo A *et al.* Nutritional factors, DNA methylation, and risk of type 2 diabetes and obesity: perspectives and challenges, *Int J Mol Sci*, 20(12), 2019, 2983.
- 67. Intine R V, Sarras M P. Metabolic memory and chronic diabetes complications: Potential role for epigenetic mechanisms, *Curr Diab Rep*, 12(5), 2012, 551-559.
- 68. Kowluru R A. Diabetic retinopathy, metabolic memory and epigenetic modifications, *Vision Res*, 139, 2017, 30-38.
- 69. Zhao S, Li T, Li J *et al.* miR-23b-3p induces the cellular metabolic memory of high glucose in diabetic retinopathy through a SIRT1-dependent signalling pathway, *Diabetologia*, 59(3), 2016, 644-654.
- 70. Gong Q, Su G. Roles of miRNAs and long noncoding RNAs in the progression of diabetic retinopathy, *Biosci Rep*, 37(6), 2017, BSR20171157.
- 71. Long J, Wang Y, Wang W, Chang B H J, Danesh F R. MicroRNA-29c is a signature MicroRNA under high glucose conditions that targets sprouty homolog 1, and its in vivo knockdown prevents progression of diabetic nephropathy, *J Biol Chem*, 286(13), 2011, 11837-11848.
- 72. Fouad M, Salem I, Elhefnawy K, Raafat N, Faisal A. MicroRNA-21 as an early marker of nephropathy in patients with type 1 diabetes, *Indian J Nephrol*, 30(1), 2020, 21-25.
- 73. Williams K T, Garrow T A, Schalinske K L. Type I diabetes leads to tissue-specific dna hypomethylation in male rats, *J Nutr*, 138(11), 2008, 2064-2069.
- 74. Williams K T, Schalinske K L. Tissue-specific alterations of methyl group metabolism with DNA hypermethylation in the Zucker (type 2) diabetic fatty rat, *Diabetes Metab Res Rev*, 28(2), 2012, 123-131.
- 75. Sifuntes-Franco S, Padilla-Tejeda D E, Carrillo-Ibarra S, Miranda-Diaz A G. Oxidative stress, apoptosis, and mitochondrial function in diabetic nephropathy, *Int J Endocrinol*, 2018, Article ID: 1875870, 2018, 13.
- 76. Santos J M, Kowluru R A. Impaired transport of mitochondrial transcription factor A (TFAM)

Available online: www.uptodateresearchpublication.com

and the metabolic memory phenomenon associated with the progression of diabetic retinopathy, *Diabetes Metab Res Rev*, 29(3), 2013, 204-213.

- 77. Mposhi A, Van Der Wijst M G P, Faber K N, Rots M G. Regulation of mitochondrial gene expression, the epigenetic enigma, *Front Biosci* - *Landmark*, 22(7), 2017, 1099-1113.
- 78. Lai C C, Shih T P, Ko W C, Tang H J, Hsueh P R. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and coronavirus disease-2019 (COVID-19): The epidemic and the challenges, *Int J Antimicrob Agents*, 55(3), 2020, 105924.
- 79. Raj K, Rohit, Ghosh A, Singh S. Coronavirus as silent killer: recent advancement to pathogenesis, therapeutic strategy and future perspectives, *Virus Disease*, 31(2), 2020, 137-145.
- 80. Pesheve E, Jiang K. Ending the pandemic, massachusetts consortium on pathogen readiness, *Harvard Medical School*, Accessed July 27, 2020.
- Balakrishnan L, Milavetz B. Epigenetic regulation of viral biological processes, *Viruses*, 9(11), 2017, 346.
- Balvan S C, García Carranca A, Song J, Recillas-Targa F. Epigenetics and animal virus infections, *Front Genet*, 6, 2015, 48.
- 83. Schafer A, Baric R. Epigenetic landscape during coronavirus infection, *Pathogens*, 6(1), 2017, 8.
- 84. Menachery V D, Schäfer A, Burnum-Johnson K E et al. MERS-CoV and H5N1 influenza virus antagonize antigen presentation by altering the epigenetic landscape, *Proc Natl Acad Sci, USA*, 115(5), 2018, E1012-E1021.
- 85. Marazzi I, Ho J S Y, Kim J *et al.* Suppression of the antiviral response by an influenza histone mimic, *Nature*, 483(7390), 2012, 428-433.
- 86. Di Mauro Gabriella, Cristina S, Concetta R, Francesco R, Annalisa C. SARS-Cov-2 infection: response of human immune system and possible implications for the rapid test and treatment, *Int Immunopharmacol*, 84, 2020, 106519.
- 87. Wrapp D, Wang N, Corbett K S *et al*. Cryo-EM structure of the 2019-ncov spike in the prefusion conformation, *bio Rxiv Prepr Serv Biol*,
- July September

367(6483), 2020, 1260-1263.

- 88. Tay M Z, Poh C M, Renia L, Mac Ary P A, Ng L F P. The trinity of COVID-19: immunity, inflammation and intervention, *Nat Rev Immunol*, 20(6), 2020, 363-374.
- 89. Shi Y, Wang Y, Shao C *et al.* COVID-19 infection: The perspectives on immune responses, *Cell Death Differ*, 27(5), 2020, 1451-1454.
- 90. Zhao J, Yuan Q, Wang H, Liu W, Liao X, Su Y *et al.* Antibody responses to SARS-CoV-2 in patients of novel coronavirus disease 2019, *Clinical Infectious Diseases*, 344, 2020, 1-22.
- 91. Long Q, Deng H, Chen J *et al.* Antibody responses to SARS-CoV-2 in COVID-19 patients: the perspective application of serological tests in clinical practice, *medRxiv*, 2019, 1-28.
- 92. Grifoni A, Weiskopf D, Ramirez S I *et al.* Targets of T cell responses to sars-cov-2 coronavirus in humans with covid-19 disease and unexposed individuals, *Cell*, 181(7), 2020, 1489.
- 93. Braun J, Loyal L, Frentsch M *et al.* Presence of SARS-CoV-2 reactive T cells in COVID-19 patients and healthy donors, *medRxiv*, 2020, 1-12.
- 94. Ferguson N M, Laydon D, Nedjati-Gilani G et al. Impact of non-pharmaceutical interventions (NPIs) to reduce COVID-19 mortality and healthcare demand, *Imperial College COVID-19 Response Team*, 2020, 1-20.
- 95. Corley M J, Ndhlovu L C. DNA methylation analysis of the COVID-19 host cell receptor, Angiotensin I converting enzyme 2 gene (ACE2) in the respiratory system reveal age and gender differences, *Preprints*, 2020.
- 96. Pinto B G G, Oliveira A E R, Singh Y *et al.* ACE2 expression is increased in the lungs of patients with comorbidities associated with severe COVID-19, *MedRxiv*, 2020, 1-16.
- 97. Zill P, Baghai T C, Schüle C *et al.* DNA methylation analysis of the angiotensin converting enzyme (ACE) gene in major depression, *PLoS One*, 7(7), 2012, e40479.
- 98. Cosma M, Staples K J, Mc Kendry R, Sanchez-Elsner T, Wilkinson T. Epigenetic control of antiviral and innate immune response following

Available online: www.uptodateresearchpublication.com

viral infection in a lung epithelial model, *European Respiratory Society Annual Congress*, 42(57), 2013, 3513.

- 99. Jin J M, Bai P, He W, Wu F, Liu X F, Han D M et al. Gender differences in patients with COVID-19: Focus on severity and mortality, *Med Rxiv*, 8, 2020, 152. Ruan S. Likelihood of survival of coronavirus disease 2019, *Lancet Infect Dis*, 20(6), 2020, 630-631.
- Leung J M, Yang C X, Tam A *et al.* ACE-2 expression in the small airway epithelia of smokers and COPD patients: Implications for COVID-19, *Eur Respir J*, 55(5), 2020, 2000688.
- 101. Bandopadhyay A R, Chatterjee D, Ghosh K, Sarkar P. COVID-19: An epigenetics and host genetics appraisal, *AJMS*, 11(3), 2020, 71-76.
- 102. Walden M, Tian L, Ross R L *et al.* Metabolic control of BRISC-SHMT2 assembly regulates immune signalling, *Nature*, 570(7760), 2019, 194-199.
- 103. Mehta P, McAuley D F, Brown M, Sanchez E, Tattersall R S, Manson J J. COVID-19: Consider cytokine storm syndromes and immunosuppression, *Lancet*, 395(10229), 2020, 1033-1034.
- 104. Sawalha A H, Zhao M, Coit P, Lu Q. Epigenetic dysregulation of ACE-2 and interferon -regulated genes might suggest increased COVID-19 susceptibility and severity in lupus patients, *Clin Immunol*, 215, 2020, 108410.
- 105. Spalluto M C, Staples K J, Mc Kendry R, Sanchez-Elsner T, Wilkinson T M A. Epigenetic control of antiviral and innate immune response following viral infection in a lung epithelial model, *Eur Respir J*, 42(57), 2013, 3513.
- 106. Hansen K D, Timp W, Bravo H C et al. Increased methylation variation in epigenetic domains across cancer types, *Nat Genet*, 43(8), 2011, 768-775.
- 107. Berman B P, Weisenberger D J, Aman J F *et al.* Regions of focal DNA hypermethylation and long-range hypomethylation in colorectal cancer coincide with nuclear laminag-associated domains, *Nat Genet*, 44(1), 2012, 40-46.
- 108. Heyn H, Vidal E, Ferreira H J *et al.* Epigenomic analysis detects aberrant superenhancer DNA methylation in human cancer,
- July September

Genome Biol, 17(1), 2016, 11.

- 109. Feinberg A P, Longo D L. The key role of epigenetics in human disease prevention and mitigation, *N Engl J Med*, 378(14), 2018, 1323-1334.
- 110. Jayaraman S. Epigenetic mechanisms of metabolic memory in diabetes, *Circ Res*, 110(8), 2012, 1039-1041.
- 111. Ihnat M A, Thorpe J E, Ceriello A. Hypothesis: The metabolic memory, the new challenge of diabetes, *Diabet Med*, 24(6), 2007, 582-586.
- 112.Illeneuve L M, Reddy M A, Lanting L L, Wang M, Meng L, Natarajan R. Epigenetic histone H3 lysine 9 methylation in metabolic memory and inflammatory phenotype of vascular smooth muscle cells in diabetes, *Proc Natl Acad Sci*, 105(26), 2008, 9047-9052.
- 113. Yu X Y, Geng Y J, Liang J L *et al.* High levels of glucose induce metabolic memory in cardiomyocyte via epigenetic histone H3 lysine 9 methylation, *Mol Biol Rep*, 39(9), 2012, 8891-8898.
- 114. Costantino S, Paneni F, Mitchell K *et al.* Hyperglycaemia-induced epigenetic changes drive persistent cardiac dysfunction via the adaptor p66 Shc, *Int J Cardiol*, 268, 2018, 179-186.
- 115. Paneni F, Volpe M, Luscher T F, Cosentino F. SIRT1, p66Shc and set7/9 in vascular hyperglycemic memory: bringing all the strands together, *Diabetes*, 62(6), 2013, 1800-1807.

- 116. Fodor A, Cozma A, Karnieli E. Personalized epigenetic management of diabetes, *Per Med*, 12(5), 2015, 497-514.
- 117. Nehme Z, Pasquereau S, Herbein G. Control of viral infections by epigenetictargeted therapy, *Clin Epigenetics*, 11(1), 2019, 55.
- 118. Lutolf M P, Gilbert P M, Blau H M. Designing materials to direct stem-cell fate, *Nature*, 462(7272), 2009, 433-441.
- 119. Roy S, Yadav S, Dasgupta T, Chawla S, Tandon R, Ghosh S. Interplay between hereditary and environmental factors to establish an *in vitro* disease model of keratoconus, *Drug Discov Today*, 24(2), 2019, 403-416.
- 120. Ghaffari L T, Starr A, Nelson A T, Sattler R. Representing diversity in the dish: Using patient-derived *in vitro* models to recreate the heterogeneity of neurological disease, *Front Neurosci*, 12(FEB), 2018, 56.

Please cite this article in press as: Aditi Munmun Sengupta *et al.* Role of epigenetics in etiology and prevention of human diseases: A review based on current evidences, *Asian Journal of Research in Biological and Pharmaceutical Sciences*, 8(3), 2020, 85-100.